ISPst9, an ISL3-like insertion sequence from Pseudomonas stutzeri AN10 involved in catabolic gene inactivation

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Received 19 March 2008 · Accepted 15 May 2008

Summary. A novel insertion sequence (IS), ISPst9, from Pseudomonas stutzeri AN10 was cloned and characterized. ISPst9 is a typical bacterial IS, consisting of a 2472-bp element flanked by 24-bp perfect inverted repeats that generates 8-bp AT-rich target duplications upon insertion. The sequence also contains a gene that encodes an active transposase (TnpA) with significant amino acid identity to members of the ISL3 family. Southern blot analysis of digested genomic DNA of strain AN10 and its 4-chlorosalicylate-degrading derivative strain AN142 demonstrated that native ISPst9 transposes in multiple copies, with one of them responsible for the nahH insertional inactivation observed in strain AN142. Precise excision of ISPst9 yielded NahH+ revertants of AN142 at high frequencies (up to 10−6). In vivo transposition, mainly in multiple copies, of an ISPst9 derivative containing a KmR cassette cloned into a suicide vector was also demonstrated. Hybridization experiments carried out with different strains of P. stutzeri and with 292 phylogenetically distinct environmental isolates suggested that the presence of an ISPst9-like IS occurs in diverse bacteria together with the presence of aromatic hydrocarbon-degrading determinants. [Int Microbiol 2008; 11(2):101-110]

Key words: Pseudomonas stutzeri · insertion sequences · mobile elements · transposition · catabolic gene inactivation

Introduction

Pseudomonas stutzeri AN10 is a naphthalene-degrading bacterium able to aerobically dissipilate naphthalene, 2-methyl-naphthalene, and salicylate as sole carbon and energy sources [44]. As described for other Pseudomonas, these aromatic hydrocarbons are funneled to the Krebs cycle through catechol (and its methyl-derivatives) [15]. Naphthalene degradation genes (nah genes) of P. stutzeri AN10 are chromosomally encoded and organized in four operons: (i) the upper pathway, coding for enzymes involved in the conversion of naphthalene to salicylate (genes nahAaAbAcAdBFCED); (ii) the lower pathway, encoding enzymes that convert salicylate to catechol and further to pyruvate and acetyl-CoA (genes nahGTHINLOMKJ); (iii) the regulatory gene nahR; and (iv) a second salicylate hydroxylase gene (nahW) [8–10]. To improve the catabolic potential of strain AN10, a derivative strain, designated P. stutzeri AN142, was constructed in our laboratory (Ginard M, 1997, Ph.D. thesis). Strain AN142 is a P. stutzeri AN10 spontaneous mutant in nahH (catechol 2,3-dioxygenase gene) that received the clc element of Pseudomonas sp. B13 carrying the genes comprising the chlorobenzoate-degradative pathway (clcABDE) [40] by conjugative transfer, thus allowing growth of the strain on 4-chlorosalicylate.

As described for many other catabolic genes [35,47], entire copies and the remnants of insertion sequence (IS) elements were found beside the nah determinants of strain...
AN10 [8–10]. IS elements are the simplest mobile genetic elements (usually less than 2.5 kb in length); they generally encode a protein required for transposition (transposase) and are found in the genomes of nearly all bacteria [11,32]. As traditionally described for the acquisition of antibiotic resistance [34], the presence of IS elements adjacent to catabolic genes has provided further support for the modular theory of evolution of modern catabolic pathways, suggesting that catabolic modules are recruited, assembled, and mobilized between bacteria by transposition events [10,52,55]. The in situ spread and even de novo construction of catabolic pathways in bacteria have allowed bacterial communities to rapidly adapt to the presence of xenobiotic organic compounds [reviewed in 50]. In addition to their role in gene recruitment and mobilization, IS elements of nearly all known IS families have been described as agents involved in the modulation of gene expression, either by polar mutation due to direct gene disruption, indirect reduction of expression of genes downstream of the insertion point, or by transcriptional activation of silent genes due to the presence of outwardly directed regulatory sequences [11,32]. Of special interest for this study are three well-characterized IS elements belonging to the ISL3 family: IS1411 from the phenol-degrading Pseudomonas sp. strain EST1001, discovered as a result of insertional activation of promotorless phenol degradation genes (pheBA) [28]; ISPst2 of Pseudomonas sp. strain OX1, formerly P. stutzeri strain OX1 [4,13,39], which inactivates the m-xylene and p-xylene catabolic pathway as well as the o-xylene catabolic genes in its derivative strain M1 [7]; and ISPpv12 from the toluene-xylene catabolic plasmid pWW0 of P. putida mt-2, whose insertional inactivation in xylE (cataloch 2,3-dioxygenase-encoding gene) allowed, after conjugative transfer of this plasmid to Pseudomonas sp. B13, the generation of derivative strain WR126, able to grow with 4-chlorobenzoate as unique carbon and energy source [56].

In this study we describe ISPst9, a novel ISL3-like IS element found in P. stutzeri strain AN10, and present results demonstrating its role as an inactivation/reactivation agent of catabolic genes, its wide distribution in different bacteria, and its occurrence together with aromatic-hydrocarbon-degrading genes.

**Materials and methods**

**Media and culture conditions.** Pseudomonas strains were grown on minimal medium [2] supplemented with either 10 mM succinate or 5 mM salicylate (or derivatives) as required. From a beach polluted after the Prestige oil spill (Galicia, Spain), 292 phylogenetically distinct environmental isolates were obtained. These isolates were grown at 25°C on minimal [2] or marine media supplemented, when necessary, with 10 mM succinate. Marine medium was prepared with artificial sea water (Scharlau) buffered with 0.1 M Tris-HCl, pH 7.4, and supplemented with 0.27 g NH₄Cl/l and 89 mg Na₂HPO₄ · 7 H₂O/l as nitrogen and phosphorus sources. An autoclaved solution of Fe(NH₄)₂(SO₄) · 6H₂O was added at a final concentration of 2 mg/l. For solid marine medium, double-strength liquid medium was prepared and mixed with an equal volume of melted, sterile, washed agar (Scharlau). Escherichia coli and Klebsiella pneumoniae strains were grown at 30°C in Luria-Bertani (LB) medium [45]. Ampicillin, kanamycin, and tetracycline were added to final concentrations of 100, 50, and 10 μg/ml, respectively, to select for the presence of plasmids. LB medium supplemented with rifampicin (40 μg/ml) was used to obtain and maintain E. coli strain DH5α RifR.

**General DNA manipulations.** Plasmids were purified using the QIAprep Spin Miniprep Kit (Qiagen). Genomic DNA from Escherichia, Klebsiella, and Pseudomonas strains were prepared by the method of Dhasee and co-workers [14]. Genomic DNA preparations from environmental isolates were done as previously described [54], with three additional freeze-thaw steps in liquid nitrogen. Restriction endonuclease digestions (Promega and GE Healthcare) and ligations with T4 DNA ligase (Invitrogen) were done as recommended by manufacturers. DNA fragments were recovered from agarose gels using the QIAquick Gel Extraction Kit (Qiagen). All other DNA manipulations were carried out according to standard procedures [45].

**PCR amplification and hybridization.** The location of ISPst9 in the nac cluster of P. stutzeri AN10 and its derivatives was routinely analyzed by PCR amplification with Taq DNA polymerase (GE Healthcare), using combinations of the following primers: ISMG4 (5'- TGGTCAGTGAC-CTTGGTC-3'), SAL23 (5'-CAAGGCCCTGAACACTGAC-3'), SAL45 (5'-GCTTCCGGTGGTGCTACGG-3'), SAL64 (5'-TCTCCGGAGTGAC-CATTGATC-3'), and SAL71 (5'-GCCACAGGCAACGGCTATTCC-3'), and the amplification conditions reported previously [36].

Southern blot hybridization was done as described by Sambrook and Russell [45]. Dot-blot hybridization (1 μg of genomic DNA per blot) was carried out with a MilliBlot-D vacuum manifold (Millipore) according to the manufacturer’s instructions. Enhanced chemiluminescence direct labeling (ECL Direct Nucleic Acid Labeling and Detection System, GE Healthcare) was used for hybridization. Gene-specific probes were prepared from P. stutzeri AN10 genomic DNA by PCR amplification with Taq DNA polymerase (GE Healthcare), using combinations of the following primers: ISMG4 (5'- ATCCAG-GCAACGGGGCTC-3') and ISMG3 (5'-CTTCCGGGCTGAGACTTGC-3') primers; nahAc probe, 0.87-kb, AC149F and AC1014R primers [19]; catA probe, 0.45-kb, 1C120F and 2C120R primers [20]; nahH probe, 0.56-kb, 3C210 and 4C230 primers [12]; nahW probe, 0.66-kb, F-nahW and R-nahW primers [9]; nosZ probe, 0.47-kb, U1672 and L2140 primers [12]. The 16S rDNA probe (1.46 kb) was generated using a mixture of genomic DNAs from different environmental isolates, the primers (16S-27F and 16S-1492R), and PCR conditions reported previously [31,36]. The kanamycin resistance gene probe was generated by excision with EcoRV of the KmR cassette from pVS2 plasmid [17]. The hybridization signal in the dots was quantified using the GeneTools analysis program (SynGene). Spearman’s rank correlation analysis of relative hybridization data was performed with SPSS 13.0 statistical data analysis package (SPSS).

**Plasmid constructions.** Plasmid pMGV01 was obtained by cloning a 4.1-kb PstI-XhoI fragment from P. stutzeri AN142 (Ginard M, 1997, Ph.D. thesis), containing the entire ISMG9, into the PstI and XhoI sites in the multicloning site (MCS) of pBluescript SK (−) (Stratagene). The fragment was obtained from P. stutzeri AN142 genomic DNA by PCR amplification with primers SAL20 (5'-GCTTCGCTCAAGGTTAAGAGG-3') and SAL25 (5'-CGGCGCAAACCTTGAATTTG-3') using the proof-reading AccuPrime PfX DNA polymerase (Invitrogen), followed by ampiclon digestion with both PstI and XhoI. Sequential restriction digestions of pMGV01 with NaeI and PvuII, followed in each case by self-ligations, yielded plasmid pJOC02. A novel PCR-generated Smal restriction site 279 bp upstream of the putative orf1 initiation codon (703-agccCGgtta-712 instead of 703-agccCGgtta-712,
coordinates from GenBank accession no. AJ582631) was constructed for PCR amplification of the entire pJOC02 plasmid with primers SMA1 (5'-AGCCGGGTTGACACAAAGC-3') and SMA2 (5'-TACCCGGGTATTGTCAGAAG-3') using AccuPrime Pfu DNA polymerase (Invitrogen), followed by restriction with SmaI and self-ligation (plasmid pJOC03). Plasmid pJOC06 was obtained by cloning the 2.76-kb EcoRI–SalI fragment from plasmid pJOC03 into the EcoRI and SalI sites in the MCS of the suicide vector pGP704 [33]. This conjugative plasmid is stable only in E. coli strains harboring the replication machinery of λ (λ strains). The EcoRI–SalI fragment was obtained by PCR amplification with primers EcoJOEC (5'-GAATTCCTGGACATGAGCAAGGCC-3') and SalJOEC (5'-GTCGACACAACCTGTTCAGCTACC-3') using AccuPrime Pfu DNA polymerase (Invitrogen). Finally, the EcoRV-excision KmR cassette from pCS12 [17] was cloned into the PCR-generated Smal site of pJOC06, generating plasmid pJOC09. E. coli strains DH5α [25] and S17.1 Δpir [26] were used as hosts in cloning experiments with pBluescript SK (−) and pGP704 derivatives, respectively.

**Phenotype reversion and transposition experiments.** For phenotype reversion experiments, P. stutzeri AN142 was grown overnight in minimal medium supplemented with 5 mM 4-chlorosalicylic acid. Culture and serial dilutions thereof were plated onto four sets of minimal medium agar plates containing 10 mM succinate (for counting all AN142 descendants), and 5 mM 3-, 4-, and 5-methylsalicylate (for counting nahH revertants), and the plates were incubated at 30°C for 48 h. The frequency of reversion was calculated from three independent experiments for each chemical compound as the ratio of revertants to all descendants. Three revertants (P. stutzeri strains R3, R4, and R5; obtained on 3-methyl-, 4-methyl-, and 5-methylsalicylate plates, respectively) were maintained for hybridization experiments.

Plasmid pJOC09 was transferred by mating with E. coli S17.1 λpir into three recipient strains: P. stutzeri AN142, a RifR spontaneous mutant of E. coli DH5α, and K. pneumoniae CMD1 [51]. Aliquots of stationary-phase cultures of donor (100 μl) and recipient (100 μl) cells were spotted together onto the surface of a LB agar plate and incubated at 30°C for 6 h. Afterwards, the cell mixture was re-suspended in 1 ml of Ringer solution (Merck). This mixture was serially diluted and plated onto minimal medium agar plates containing succinate plus kanamycin (for transconjugants), or succinate alone (for all recipients), and the plates were incubated at 30°C for 48 h. The frequency of conjugation plus transposition for each strain was calculated from three independent experiments as the ratio of transconjugants to all recipients.

**DNA sequencing and analysis.** Nucleotide sequences were determined (both strands) directly from plasmids pMGV01 and pPS05-C [10] using the BigDye terminator cycle sequencing kit (Applied Biosystems) according to the manufacturer’s instructions. Sequences were extended by primer-walking, with the design of new primers based on the sequences determined in this study. Primer design and sequence analyses were done using BioEdit 6.0.5 sequence alignment editor [24], Vector NTI 10.0 suite (Invitrogen), and EMBOSS suite [42]. Similarity searches with GenBank and EMBL databases were done using BLASTP (NCBI) and FASTA-protein (EBI) web tools, respectively [1,38]. The ClustalX program [48] was used for amino acid sequence alignments. The Protidist program of the PHYLIP 3.6 package [18] and PhyML program [22] were used for inferring distance- and likelihood-based phylogenies, respectively. Kimura’s distance, Dayhoff PAM matrix, and the Jones-Taylor-Thornton model were used as models of amino acid substitution in both approaches. Bootstrap (100 replicates) was undertaken to construct distance-based trees using the neighbor-joining method. Resulting phylogenies were displayed using the TreeView 1.6.6 program [37].

**Nucleotide sequences accession numbers.** The sequence of ISPst9, as determined from the insert in pMGV01, was deposited in GenBank under accession no. AJ582631. The GenBank accession no. for the sequence of the P. stutzeri AN10 ISPr9 insertion sequence and its flanking region is DQ473406.

**Results**

**Isolation and characterization of ISPst9.** ISPr9 was discovered during an analysis of the nahH (catechol 2,3-dioxygenase-encoding gene) mutation present in P. stutzeri AN142, the 4-chlorosalicylate-degrading derivative of P. stutzeri AN1. A PstI–Xhol-digested chromosomal DNA of AN142 and the parental P. stutzeri strain AN10 were subjected to electrophoresis and Southern blot hybridization against a nahH probe generated by PCR from strain AN10. The results indicated that P. stutzeri AN142 had acquired a 2.5-kb DNA insertion in nahH (Fig. 1A). After PCR amplification of the insert with primers SAL20 and SAL25 (a schematic representation of their location on P. stutzeri AN142 genome is shown in Fig. 1B) and cloning of the resulting fragment to generate plasmid pMGV01, the complete nucleotide sequence of the DNA insertion on nahH in P. stutzeri AN142 was determined. Sequence analysis revealed that the DNA insertion was 2472 bp in length and occurred after nucleotide 159 of nahH. The DNA insertion was flanked by an 8-bp direct repeat of nahH bases 152–159 (5′-ACAAAAATCC-3′). It had two perfect 24-bp inverted repeats at either end (5′-GGGT-ATGCCTGGATTATAATGGTGAT-3′) and coded for a putative transposase (see below). Thus, as it contained the main features of an insertion sequence, the insert was named ISPst9, accordingly to the IS database [http://www-is.biotoul.fr/is.html].

Computer-aided analysis of the ISPst9 DNA sequence indicated the presence of two potential open-reading frames (ORFs) transcribed in the same direction (Fig. 1B). The first ORF (orf1) coded for a putative protein of 206 amino acids with a predicted mass of 21.8 kDa. The amino acid sequence analysis indicated that this protein belongs to a LysE-type translocase family (pfam01810) as it showed the highest amino acid identity values (41–43%) with LysE-like proteins from whole-genome sequences of Pseudomonas species. The second ORF (tnpA4) coded for a putative protein of 429 amino acids with a predicted mass of 49.3 kDa. The deduced amino acid sequence of tnpA4 (TnpA4) showed the highest level of homology with two transposase-like putative proteins annotated from the whole-genome sequence of Idiomarina loihiensis (Fasta amino acid identity: 93 and 92%, in 429 amino acids, with Q5X23 and Q5UL5, respectively) [27]. In addition, TnpA4 is highly homologous to two well-characterized ISL3-family transposases from hydrocarbon-degrading Pseudomonas strains (FASTA amino acid identity: 93% in 404 amino acids with OS2212, the ISPpsu1 transposase from P. putida mt-2, and 87% in 423 amino acids with Q9X7J2, the ISPst2 transposase from Pseudomonas sp. OX1) [7,53,56]. Multiple alignments based on these and other ISL3-like transposases retrieved from the IS database were conducted. The results showed that the
sequence of TnpA4 had the conserved DDE motif of active transposases [11]. Phylogenetic trees were generated for TnpA4 and homologous transposases retrieved from the IS database (> 40% amino acid identity). All distance-based and maximum-likelihood phylogenetic approaches showed, independent of the amino acid substitution model used, that the sequence of IS\textit{Pst} \textit{9} transposase TnpA4 grouped with IS\textit{L} \textit{3} transposases from environmental gamma-proteobacterial isolates, mainly aromatic-hydrocarbon-degrading bacteria (Fig. 2, group 1), and was clearly distinct from those in group 2, mainly constituted by IS\textit{L} \textit{3}-like insertion sequences identified in clinical isolates.

**Location and number of IS\textit{Pst} \textit{9} in the genome of \textit{P. stutzeri} AN10 and its derivative \textit{P. stutzeri} AN142.** Sequence analysis of IS\textit{Pst} \textit{9} from \textit{P. stutzeri} AN142 revealed that 94 nucleotides at its 3'-end, downstream of \textit{tnpA4}, overlapped with 100% identity with the 5'-end of the sequence of a genome fragment from \textit{P. stutzeri} AN10, which contains the regulatory- (\textit{nahR}) and catabolic-salicylate-degrading (\textit{nahW} and \textit{nahGTHINLOMKJ}) genes as determined previously (sequence AF039534 [9,10]). This result suggested that a copy of IS\textit{Pst} \textit{9} was originally located beside salicylate-degrading genes in strain AN10. To prove this, we sequenced the nucleotides located upstream \textit{tnpA3} (an IS\textit{5}-like transposase-encoding gene) of \textit{P. stutzeri} AN10, which was already known [10]. Sequence analysis confirmed the presence in \textit{P. stutzeri} AN10 of an identical copy of IS\textit{Pst} \textit{9} (100% nucleotide identity) upstream of \textit{tnpA3} (Fig. 3A). Computer-aided analysis also showed the presence of two partial (‘\textit{orf2} and \textit{orf4}’) and one complete ORF (\textit{orf3}) located upstream of IS\textit{Pst} \textit{9} in \textit{P. stutzeri} AN10 (Fig. 3A). The ‘\textit{orf2}’ partial C-terminal gene product (153 amino acids) was homologous to the C-terminal domain of acetoacetyl-CoA synthases from several \textit{Pseudomonas} strains (amino acid identity values: 73.8% to Q9I2B2 of \textit{P. aeruginosa} PAO1, 75.7% to Q4K4Q4 of \textit{P. fluorescens} Pf-5, and 71.0% to Q88IC8 of \textit{P. putida} KT2440). The \textit{orf3} gene product (330 amino acids) resembled a lactone hydrolase (62.4% amino acid identity to the CamQ lactone hydrolase of pCAM plasmid from \textit{P. putida} NCIMB 10007, Q6STL9). Finally, the \textit{orf4’} partial C-terminal gene product (96 amino acids) showed 62.8% identity to the C-terminal domain of the CamR transcriptional repressor encoded on pCAM plasmid from \textit{P. putida} PpG1, Q6STL9 [3].

To determine whether the original copy of IS\textit{Pst} \textit{9} present in \textit{P. stutzeri} AN10 was maintained in \textit{P. stutzeri} AN142 after the transposition event, shown in Fig. 1A, a set of PCR reactions was developed to analyze the location of IS\textit{Pst} \textit{9} in \textit{P. stutzeri} AN142. The presence of IS\textit{Pst} \textit{9} upstream of \textit{tnpA3} was evaluated with primers SAL71, SAL64, and ISMG4 (a schematic representation of their location is shown in Fig. 3A); inactivation of \textit{nahH} by IS\textit{Pst} \textit{9} in \textit{P. stutzeri} AN142 was confirmed using primers SAL23, SAL45, and ISMG4 (a schematic representation of their location is shown in Fig. 1B). As shown in Fig. 3B, PCR amplifications using the

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**Fig. 1. Location of IS\textit{Pst} \textit{9} in \textit{Pseudomonas stutzeri} strain AN142. (A) Southern blot of \textit{P. stutzeri} AN142 and AN10 genomic DNAs digested with \textit{PstI} and \textit{XhoI} and probed with the \textit{nahH} probe. (B) Schematic representation of IS\textit{Pst} \textit{9} in \textit{P. stutzeri} AN142 and its flanking regions. Genes code for the following proteins: \textit{nahG}, salicylate hydroxylase; \textit{nahH}, catechol 2,3-dioxygenase; \textit{nahl}, hydroxymuconic semialdehyde dehydrogenase; \textit{nahI}, ferredoxin-like protein; \textit{orf1}, LysE-like protein; \textit{tnpA4}, transposase-like protein. Arrows indicate the directions of gene transcriptions. ISMG4, SAL20, SAL23, SAL25, and SAL45 denote the primers used. Thin lines indicate their location and 5'-3' orientation. IR and DR denote inverted and direct repeats of IS\textit{Pst} \textit{9}, respectively. Line below AJ582631 accession number indicates sequence data submitted to GenBank.
SAL71 and SAL64 primers and the SAL71 and ISMG4 primers produced single bands of 3.5-kb and 0.8-kb, respectively, for strains AN10 and AN142. Thus, the original copy of IS_Pst9 in _P. stutzeri_ AN10 was maintained upstream of _tnpA3_ in _P. stutzeri_ AN142. PCR amplifications with the SAL23 and SAL45 primers and the SAL23 and ISMG4 primers corroborated that _nahH_ of _P. stutzeri_ AN142 was inactivated by IS_Pst9 but was intact in _P. stutzeri_ AN10. To analyze whether more than two copies of IS_Pst9 were present in the _P. stutzeri_ AN142 genome, Southern blot hybridizations against _EcoRI-digested genomic DNAs_ from strains AN10 and AN142 were done using probes against IS_Pst9 and _nahH_ genes. The results revealed the presence of a unique copy of IS_Pst9 in _P. stutzeri_ AN10 while strain AN142 had, at least, eight copies of IS_Pst9, including the original one (Fig. 3C). Southern blot hybridization also confirmed the presence of a unique copy of _nahH_ in strain AN142, which hybridized also with a copy previously detected for IS_Pst9 (indicated with an arrow in Fig. 3C). No hybridization signal was observed in _Pseudomonas_ sp. B13 (result not shown), thus suggesting that all new IS_Pst9 copies observed in strain AN142 came from the one detected in _P. stutzeri_ AN10.

**Reversion of _P. stutzeri_ AN142 to a NahH+ phenotype.** Although _P. stutzeri_ AN142 was able to grow on 4-chlorosalicylate because it carried the _clc_ element from _Pseudomonas_ sp. B13 [40], this strain was unable to grow on methylsalicylates. This is because ortho-cleavage of methylcatechols by the catechol 1,2-dioxygenase encoded in plasmid B13 yields methylmuconolactones [49], which are dead-end metabolites. Thus, growth of strain AN142 on methylsalicylates was only possible through the recovery of methylcatechols meta-cleavage activity mediated by NahH (catechol 2,3-dioxygenase); this would imply a precise excision of IS_Pst9 from the _nahH_ gene of _P. stutzeri_ AN142. Putative NahH+ revertants able to use three different methylsalicylates were isolated by plating cultures of strain AN142, grown on 4-chlorosalicylate, on minimal medium supplemented with 3-, 4-, or 5-methylsalicylate as unique carbon and energy sources. Similar reversion frequencies were observed in all cases (1.4 ± 0.4 × 10^-6 on 3-methylsalicylate, 8.8 ± 0.6 × 10^-6 on 4-methylsalicylate, and 4.6 ± 0.5 × 10^-6 on 5-methylsalicylate). Three revertants (designated as R3, R4, and R5; from 3-, 4-, and 5-methylsalicylate plates, respectively), from each of the isolation strategies, were selected for further analysis. PCR analysis showed that all revertants maintained the IS_Pst9 copy upstream of _tnpA3_, the original position in AN10, and that they had lost the IS_Pst9 copy interrupting _nahH_ (results not shown). Furthermore, Southern blot hybridizations against _EcoRI-digested genomic DNAs_ from all three revertants confirmed the loss of only one copy of IS_Pst9, that interrupted _nahH_ (indicated with an arrow in Fig. 3C), and suggested the recovery of the original _nahH_ gene structure. To clarify how the excision of IS_Pst9 restored...
NahH function, the nahTH region of all three revertants (R3, R4, and R5) was amplified using SAL23 and SAL45 primers (a schematic representation of their location is showed in Fig. 1B) and sequenced. The nahH sequences of all three revertants were identical to that of strain AN10, not conserving the 5′-ACAAATTC-3′ direct repeat sequence duplicated in the insertion event in strain AN142. This result confirmed the precise excision of ISPst9 from nahH.

**Demonstration of ISPst9 transposition in other bacteria.** To demonstrate that ISPst9 was able to transpose, plasmid pJOC09, carrying ISPst9 with an inserted KmR cassette as phenotypic marker, was constructed as described in Materials and methods. In pJOC09, the KmR cassette was inserted in a PCR-generated SmaI restriction site located 279 bp upstream of the ATG codon of orf1 in ISPst9, as it was thought that, in this location, the cassette would not disable any transposition function by affecting the expression of the ISPst9 genes. As pJOC09 plasmid is a derivative of the suicide vector pGP704 [33], this plasmid is stable only in λpir derivatives of *E. coli* and can be transferred from those strains into recipient strains but will not replicate within them. Thus, conjugation experiments between *E. coli* S17.1λpir carrying pJOC09 with strains of distinct bacterial species (*K. pneumoniae* CMD1, *E. coli* DH5α RifR, and *P. stutzeri* AN10) were done. In all mating experiments, a significant number of KmR transconjugants with a wide range of transposition frequencies were obtained (9.6 ± 1.1 × 10⁻⁵ for *P. stutzeri*, 9.3 ± 0.9 × 10⁻⁶ for *E. coli*, and 7.8 ± 6.2 × 10⁻⁸ for *K. pneumoniae*). Southern blot hybridization of chromosomal digests of transconjugants...
against a Km<sup>R</sup> cassette probe revealed that most isolates harbored more than one copy of the IS<sub>Pst</sub><sup>9</sup> derivatives, as expected, with six being the highest number of copies detected (results not shown).

**ISP<sub>Pst</sub><sup>9</sup> in genomes of *P. stutzeri* strains and other bacteria.** As noted above, ISP<sub>Pst</sub><sup>9</sup> belongs to a subfamily of ISL3-like insertion sequences found mainly in aromatic-hydrocarbon-degrading bacteria (group 1 in Fig. 2): therefore, their presence in a genome may be related to the occurrence of genes encoding proteins with this function. To evaluate the simultaneous occurrence of both ISP<sub>Pst</sub><sup>9</sup> and nah genes, Southern blot hybridizations against EcoRI-digested genomic DNAs from several naphthalene-degrading (Nah<sup>+</sup>) and non-degrading (Nah<sup>-</sup>) strains of *Pseudomonas stutzeri* that belong to different genomovars (gv.). Strain sources are as follows: ATCC17587, ATCC17589, and CCUG11256<sup>T</sup> [46]; 19SMN4, AN10, DNSP21, DSM50227, and DSM50238 [43]; AN11, LSMN2, S1MN1, ST27MN2, and ST27MN3 [21]. *P. balearica* strain SP1402<sup>T</sup> (formerly *P. stutzeri*, PhSP1402) was also included [5]. Arrows indicate those EcoRI DNA fragments that hybridized with the nah<sub>W</sub> probe.

To extend this study to other bacteria, 292 environmental isolates obtained from samples polluted by *Prestige* tanker fuel that were collected at a beach in Galicia, NE Spain, and from non-polluted samples collected at the same beach were used. The phylogenetic affiliations of the isolates were determined by 16S rDNA analyses (data not shown), which revealed that 57% belonged to gammaproteobacteria (167 isolates). Of these, 33% were identified as *Pseudomonas* (97 isolates) and 13% as *P. stutzeri* (39 isolates). Dot-blot hybridizations against genomic DNA were done using DNA probes for ISP<sub>Pst</sub><sup>9</sup> and five other selected genes: 16S rDNA, used as the control gene; nah<sub>Ac</sub> (naphthalene 1,2-dioxynase α-subunit) and nah<sub>H</sub> (catechol 2,3-dioxygenase), two widespread naphthalene-catabolizing genes present in *Pseudomonas* and non-*Pseudomonas* naphthalene-degrading strains [10,23, 29]; and nos<sub>Z</sub> (nitrous oxide reductase) and cat<sub>A</sub> (catechol 1,2-dioxygenase), two genes considered as characteristic for *P. stutzeri* species and used as phylogenetic markers for this species [30]. Hybridization experiments were followed by correlation analysis of the relative

![Fig. 4](https://example.com/fig4.png)

**Fig. 4.** Southern blot hybridization with the ISP<sub>Pst</sub><sup>9</sup> probe of EcoRI-digested genomic DNAs from several naphthalene-degrading (Nah<sup>+</sup>) and non-degrading (Nah<sup>-</sup>) strains of *Pseudomonas stutzeri* that belong to different genomovars (gv.). Strain sources are as follows: ATCC17587, ATCC17589, and CCUG11256<sup>T</sup> [46]; 19SMN4, AN10, DNSP21, DSM50227, and DSM50238 [43]; AN11, LSMN2, S1MN1, ST27MN2, and ST27MN3 [21]. *P. balearica* strain SP1402<sup>T</sup> (formerly *P. stutzeri*, PhSP1402) was also included [5]. Arrows indicate those EcoRI DNA fragments that hybridized with the nah<sub>W</sub> probe.
hybridization intensities for the different genes analyzed (Table 1). As expected, high correlation values \((r = 0.84–0.88)\) were observed in the presence of both naphthalene degradation genes \((nahAc\) and \(nahH)\), independent of the phylogenetic group analyzed. The correlation between the hybridization values obtained for ISP9 and 16S rDNA probes was lower \((r = 0.37–0.62)\), as expected. Note that the correlation values between the presence of both nosZ and catA genes were highest only when \(P.\) \(stutzeri\) isolates were considered \((r = 0.79)\). The values decreased when isolates from other species were added to the analysis \((r = 0.56–0.69)\), confirming the importance of these two genes in the definition of \(P.\) \(stutzeri\) species. Correlation values between ISP9 and the naphthalene-catabolizing genes \(nahAc\) and \(nahH\) were high \((nahH, r = 0.64–0.80; nahAc, r = 0.75–0.87)\), i.e., similar to those measured for the genes thought to coexist, such as \(nahAc\) and \(nahH\).

**Table 1.** Spearman’s rank correlation \((r)\) of hybridization data obtained with environmental isolates*

<table>
<thead>
<tr>
<th>Probes used</th>
<th>All ((n = 292))</th>
<th>Gammaproteobacteria ((n = 169))</th>
<th>Pseudomonas sp. ((n = 97))</th>
<th>Pseudomonas (stutzeri) ((n = 39))</th>
</tr>
</thead>
<tbody>
<tr>
<td>ISP9 (\text{vs}) nahH</td>
<td>0.75</td>
<td>0.77</td>
<td>0.64</td>
<td>0.80</td>
</tr>
<tr>
<td>ISP9 (\text{vs}) nahAc</td>
<td>0.78</td>
<td>0.76</td>
<td>0.75</td>
<td>0.87</td>
</tr>
<tr>
<td>ISP9 (\text{vs}) 16S rDNA</td>
<td>0.37</td>
<td>0.32</td>
<td>0.62</td>
<td>0.39*</td>
</tr>
<tr>
<td>nahAc (\text{vs}) nahH</td>
<td>0.84</td>
<td>0.85</td>
<td>0.88</td>
<td>0.88</td>
</tr>
<tr>
<td>nosZ (\text{vs}) catA</td>
<td>0.63</td>
<td>0.69</td>
<td>0.56</td>
<td>0.79</td>
</tr>
</tbody>
</table>

\(P\) for each \(r\) value is \(<0.001\), with the exception of ISP9 \(\text{vs}\) 16S rDNA in \(P.\) \(stutzeri\), whose \(P\) is 0.014.

**Discussion**

This report describes a novel IS element of \(P.\) \(stutzeri\), ISP9, located adjacent to the naphthalene lower-degradation pathway of strain AN10. Phylogenetic comparisons of the ISP9 transposase (TnpA4) with its closer relatives showed that ISP9 belongs to a subfamily of ISL3-like ISs that are widespread mainly in environmental *Proteobacteria* and that it is clearly distinct from other closely related relatives found in clinical isolates. Nearly all members of the ISP9 subfamily of ISL3-like ISs are present in aromatic-hydrocarbon-degrading bacteria. Two of them, ISP2 from *Pseudomonas* sp. OX1 [7] and ISP12 of *P. putida* KT2440 [56], are involved in catabolic gene inactivation. This is also the case of ISP9, which was first detected as a DNA insertion in \(nahH\) of \(P.\) \(stutzeri\) AN142, a 4-chlorosalicylate-degrading AN10-derivative strain constructed in our laboratory (Ginard M., 1997, Ph.D. thesis). The insertion in \(nahH\) inactivates meta-cleavage of 4-chlorocatechol. This step is a biochemical prerequisite to the proper channeling of 4-chlorocatechol to intermediates of the tricarboxylic acid cycle through an ortho-cleavage pathway [41]. Note that exactly the same genotype (i.e., inactivation of the catechol 2,3-dioxygenase gene \(xylE\) by ISP9-like ISPPu12) has been reported to occur in *Pseudomonas* sp. WR26 [56]. Similar behavior was also observed for ISP2 in *Pseudomonas* sp. OX1 and in its derivative *Pseudomonas* sp. M1 [7]. In strain OX1, one copy of ISP9 is situated upstream of \(xylMA\) (xylene-monoxygenase-encoding genes), between \(xylW\) (unknown function) and \(xylC\) (benzaldehyde-dehydrogenase-encoding gene), which prevents the growth of this strain on \(-\)-xylene and \(-\)-xylene by a polar effect on \(xylUWCMABN\) expression [7]. Alternatively, strain OX1 is able to grow on \(-\)-xylene as unique carbon and energy source using the \(touABCDE\) (toluene monoxygenase operon) gene products [6]. The \(-\)-xylene and \(-\)-xylene cometabolism by toluene monoxygenase gen in strain OX1 generates unproductive-growth intermediates [6]. *Pseudomonas* sp. M1 had a recovered \(xylUWCMABN\) genotype and showed an ISP2 insertion in the \(touA\) gene of the toluene monoxygenase operon, allowing its growth on \(-\)-xylene but not on \(-\)-xylene. In addition, the original ISP9 copy of *P. stutzeri* AN10, located beside gene \(nahW\), encoding a salicylate-degrading ability, disrupted a gene plausibly encoding CamR, the transcriptional repressor of the operon for camphor catabolism. Sequence evidence obtained in this study suggested that *P. stutzeri* AN10 harbors at least part of the genetic determinants for camphor degradation, although this strain does not grow on camphor. Thus, it could be that these genes encoding camphor degradation are a remnant of a former catabolic capability of *P. stutzeri* AN10, which might have been misregulated by the insertion of ISP9 in the camR gene.

In ISP9, ISP12, and ISP2, the ISs showed the ability to transpose in multiple copies and, at least in the case of the disrupted catabolic genes, an 8-bp direct repeat was always generated [7,56]. Moreover, M1 revertants, in which ISP2 was precisely excised from the \(touABCDE\) operon,
were obtained at high frequencies (10^{-4} to 10^{-3}) when o-xylene was used as unique carbon and energy source [7,16]. ISP\textsubscript{Pu}9 had exactly the same behavior, although at lower frequencies (10^{-5}), when P. stutzeri AN142 was grown on methylsalicylates as unique carbon and energy sources. In both cases (ISP\textsubscript{Pu}9 and ISP\textsubscript{Pu}2), only the IS copy that inactivated the catabolic gene was lost in the revertants, but there were no other changes in the IS patterns. It is difficult to explain how transposition could be the only mechanism responsible for the precise excision observed in all of the revertants analyzed. For example, it is also plausible that a recombination event between the transposition-generated direct repeats resulted in the regeneration of the disrupted catabolic genes. Thus, members of the ISP\textsubscript{Pu}9 subfamily of IS\textsubscript{L3}-like ISs could act, by transposition, as inactivation agents of catabolic gene expression. Furthermore, ISP\textsubscript{Pu}9-like ISs might be precisely excised, probably helped by the recombination machinery, thus allowing the synthesis of active catabolic enzymes as needed in the respective environments. In any case, as indicated by the existence of IS\textsubscript{1411} [28], which is involved in transcriptional activation of genes encoding phenol degradation in P. putida EST1001, other IS\textsubscript{L3}-like ISs, not belonging to the ISP\textsubscript{Pu}9 subfamily, are involved in catabolic gene regulation.

Using a suicide vector, we demonstrated that ISP\textsubscript{Pu}9 transposes at high frequencies (up to 10^{-5} Kn\textsuperscript{K} transconjugants per donor) and in multiple copies, independent of the genetic background (P. stutzeri, E. coli, and K. pneumoniae). These results are similar to those obtained for ISP\textsubscript{Pu}u12 of P. putida mt-2 [56], one of its closest well-studied relatives. It was demonstrated that two identical copies of ISP\textsubscript{Pu}u12 are involved in mobilization by transposition of the dehalogenase genetic determinants (\textit{deh\textsubscript{L}} and its cognate regulatory gene, \textit{deh\textsubscript{R}}) in the DEH element in P. putida PP3 [53]. Thus, experimental and sequence evidence indicates that members of the ISP\textsubscript{Pu}9 subfamily of IS\textsubscript{L3}-like ISs are also involved in mobilizing catabolic determinants.

Both events, transcriptional regulation and mobilization of catabolic genes, suggest that ISP\textsubscript{Pu}9-like ISs and catabolic genes occur together in the same genome. Southern blot hybridization using an internal ISP\textsubscript{Pu}9-probe against EcoRI-digested genomic DNA of several P. stutzeri isolates revealed that the occurrence in this species of ISP\textsubscript{Pu}9-like ISs was not strictly dependent but more frequent in the presence of genes encoding naphthalene degradation. In fact, at least one copy of ISP\textsubscript{Pu}9, if present in a naphthalene-degrading P. stutzeri strain, could be located within the same EcoRI-EcoRI DNA fragment as \textit{nahW} (encoding salicylate hydroxylase) in P. stutzeri AN10. Furthermore, dot-blot hybridization analyses done in this study with 292 phylogenetically distinct environmental isolates revealed statistically significant correlations between the occurrence of both ISP\textsubscript{Pu}9 and the analyzed catabolic genes, regardless of the phylogenetic affiliation of the isolates. Our results strongly suggest that ISP\textsubscript{Pu}9-like ISs are widespread in bacteria, not only in \textit{Gammaproteobacteria}, as previously suggested by sequence evidence [56]; and that, statistically, their presence occurs together with that of genes encoding aromatic hydrocarbon degradation or, at least, the \textit{nah} determinants thereof.

**Acknowledgements.** We thank Margarita Ginard for providing strain AN142 and for helpful discussions. The support of the Scientific-Technical Service of the University of Balearic Islands during the operation of the genetic analyzer for sequencing is also acknowledged. BN was supported by a contract of the Ramón y Cajal program from the MEC (Spanish Ministry of Education and Science). MPL was supported by a doctoral grant of the A\'nàn Office. JAC-O and CM-C were supported by grants of the CAIB (Government of Balearic Islands). Funds were obtained from projects VEM2003-20565 and CTM2005-01783 from MEC, and project PJB2004-10152 from CAIB.

**References**


